

Single Cell RNA-Seq Analysis in Partek® Flow®

HANDS-ON TRAINING

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Experiment Description

- 1k peripheral blood mononuclear cells (PBMCs) from a healthy donor
 - Any peripheral blood cell having a round nucleus
- Partek Flow supports file types: bcl, fastq, bam, h5, txt etc
- Choose **filtered** feature/ cell matrix in .h5 format from CellRanger output
- *Goal for today: Identify different blood cell populations*

Download in browser		Batch download
If the file size is large, we suggest using batch download instead.		
Input Files	Size	md5sum
FASTQs	29.3 GB	40274800f18c380b9bd7c25e3ed43419
Output Files	Size	md5sum
Genome-aligned BAM	23.3 GB	cd02b972a841487b09782f3b1724e42e
Genome-aligned BAM index	10.5 MB	9fbb1a8593a9421e1b5320aea76e7157
Per-molecule read information	257 MB	f592a76eba137b1ad07b845628a60112
Feature / cell matrix HDF5 (filtered)	18.1 MB	4dcd7861f61219bd5325190e4c5f6798
Feature / cell matrix (filtered)	43.1 MB	f741a636ede503cf65491320ed3ec719
Feature / cell matrix HDF5 (raw)	153 MB	7c5d7164c8e8a36fa3f6e334fb0281cd
Feature / cell matrix (raw)	89.6 MB	6616dcd7c6c5275b66ff7d6b806002c7
Clustering analysis	27.1 MB	a96940bfa361d8b52953047bb1afca79
Summary CSV	683 B	6178a3305959996189a48f3d5d73bb82
Summary HTML	3.95 MB	7c386237483575edccd1362e14ff7027
Loupe Browser file	65.1 MB	370d0bc8472a161054ac5d6209e4de56

Import data

Login

- Open a web browser (Chrome, Firefox, Safari etc.)
- Go to the server URL
- Log in with your username and password
- This will open to the Partek Flow homepage

Create project

- Click **New Project** and enter project name
- Click **Add data**
- Select scRNA-seq>10X Genomics Cell Ranger count h5
- Click **Next**
- User file browser to select file
- Select feature annotation file
- Click **Finish**

Single Cell QA/QC

Run QA/QC task

- Click the **Single cell counts** data node
- Select **Single Cell QA/QC** from the **QA/QC** section of the task menu

View report and filter cells

- Double click on **Single cell QA/QC** report to open it
- Click **Select & Filter** in Tools section to specify filter criteria
- Set Detected features **min** to **200**
- Set %Mitochondrial counts **max** to **15**
- Click **Filter include**
- Click **Apply observation filter**
- Choose the **Single cell counts** data node and click **Select**
- Click **Save** in the data viewer to save the filter session

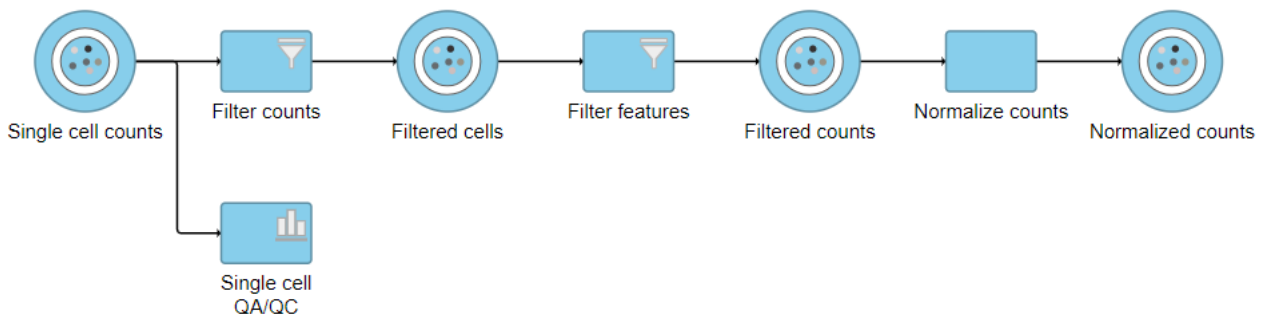
Filter Features & Normalization

Filter genes

- Click **Filter features** in Filtering section on the menu
- Choose **Noise reduction**
- Filter criteria: Exclude **value ≤ 0 in at least 99%** of the cells
- Click **Finish**

Normalization

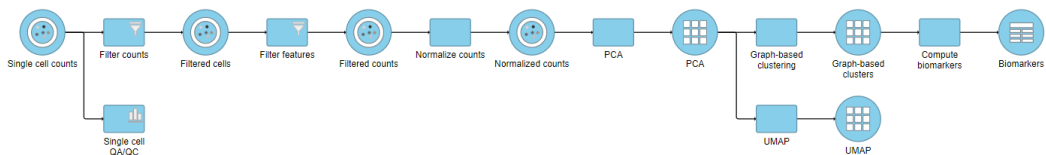
- Click on **Filtered count node > Normalization in Normalization and scaling** section on the menu
- Click **Use recommended**
- Click **Finish**



Exploratory analysis

Dimension reduction

- Click the **Normalized counts data** node
- Click **PCA** in the **Exploratory analysis** section of the task menu
- Click **Finish** to run with default settings
- Double click on **PCA** node to view report
- Select **PCA** node, choose **UMAP** task on the menu
- Click **Finish** with default settings
- Select **PCA** node, choose **Graph-based clustering** task
- Click **Finish** with default settings
- Double click on **Graph-based clusters** node to view the report
- Double click on **Biomarkers** node to view the report



View Gene Expression in UMAP



- Double click **UMAP** to open the report
- Click **Get data**, search for **Graph-based clusters** node, click on the node, choose **Graph-based** annotation, drag and drop to **Color UMAP**
- From **Get data** dialog, search for **Biomarkers** data node, drag the node and drop to the **Bottom** of **UMAP** plot to add the biomarker table in the viewer
- Click **Duplicate plot** in UMAP plot to make a copy of UMAP
- Drag a gene from the biomarker table to drop on one of the UMAP plot to color the cells
 - Use one gene to color cells
 - Use two or three genes to color cells
 - Use gene list to color cells

Cell Type	Gene Markers
T-cells	CD3D, CD3E
Cytotoxic cells	NKG7, GNLY
B cells	CD79A, CD79B (list)
Monocytes	CD68, CD14

Classify Cell Types

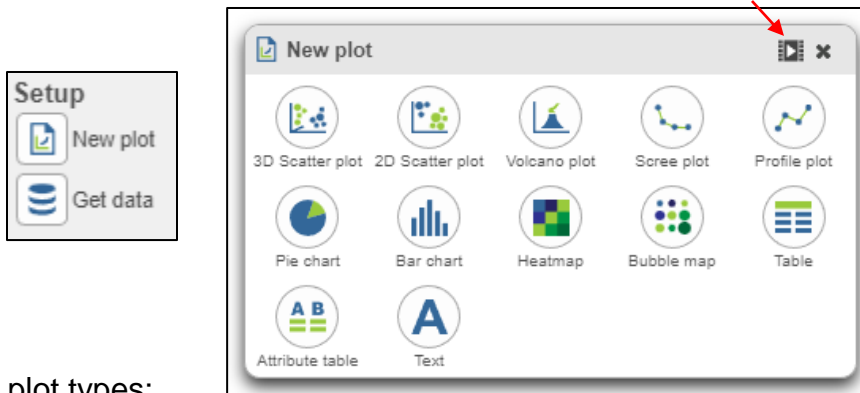
- Click on **Select & Filter** in Tools menu
- Drag and drop legend from plots to add criteria
 - Use graph-based annotation
 - Use gene expression
- Search attributes or gene to add criteria
 - Click on the circle next to the criteria drop-down box to select correct data node
 - No limitation on the number of rules
 - And operation is performed among multiple rules
 - Cells meet the criteria will be selected and highlighted in the plots
- Once cells are selected, click on **Classify** in Tools menu, choose **Classify selection** to give a name to the selected cells
 - Always **Save** the analysis
 - Click on **Style > Color by > New classification** to preview the cell types before apply classification
 - Only click **Apply classifications** when all cell types are identified
- Once classification is applied, it will appear in Meta data tab. This cell attribute is project level, which means all data nodes in the project and see it.

Identifying Differentially Expressed Genes

- Click the **Normalized counts** node
- Click **Statistics** then the **Differential analysis** section of the task menu
- Choose **ANOVA** as the method and click **Next**
- Choose **Cell type** and click **Add factors**
- Click **Next**
- Choose to compare **B cell** vs **T cell**, click **Add comparison**
- Click **Finish**
- Double click the **ANOVA** data node to open the ANOVA report
- Click the  icon next to a gene under **View** to open data viewer
- Using the **Style** configuration option under *Summary*, add violins or box Whiskers on the plot
 - Duplicate the plot and change Y-axis to view multiple genes
 - Drag and drop group name on X-axis to change order
- Click the  icon to invoke volcano plot
 - Use on **Style** > **Label by** to display selected gene name
 - Use **Select & Filter** to search for genes
- Use Gene list filter panel on ANOVA report to generate gene list based on specified criteria

Plot Types

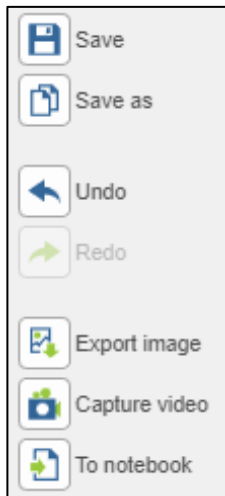
- Use **New plot** or **Get data** under *Setup* to add content to your *Data Viewer* session
- **Tip: watch the help videos to navigate each of the panel options**



- The plot types:
 - *Text*. Inserts a text box. Useful for captions and notes
 - *Table*. Inserts a table. E.g. table of biomarkers per cluster
 - *Heatmap*. Inserts a heat map, e.g. result of hierarchical clustering
 - *Bubble map*. Uses 3 dimensions to display groups by summary statistics resulting from the hierarchical clustering/ heatmap task
 - *3D Scatter Plot*. Inserts a 3D scatterplot. This is a plot showing three data features at the same time.
 - *2D Scatter Plot*. Inserts a 2D scatter plot. This is a plot showing two data features at the same time.
 - *Volcano plot*. Visualize significance and the magnitude of changes in features within a comparison
 - *Scree plot*. This plot shows information content per principal component.
 - *Attribute summary table*. Insert a table which shows attributes
 - *Histogram*. Histogram of a variable
 - *Profile plot*. Bar or line chart on one or multiple variables
 - *Pie chart*. Inserts a pie chart. Useful for visualization of categorical variables.

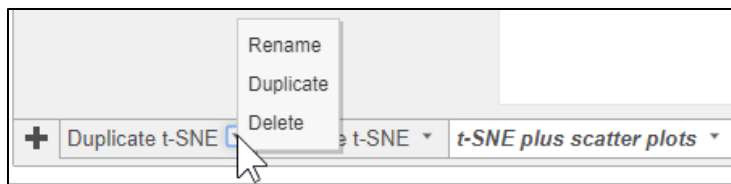
General Data Viewer Controls

- General control buttons are on the left






- **Save.** Saves the session to the *Data Viewer* tab.
- **Save As...** Saves the session under a new name
- **Undo** (*right-click to see/select multiple steps*)
- **Redo** (*right-click to see/select multiple steps*)
- **Export Image.** Saves the entire canvas as an image to the local computer. Supported formats are .png, .svg and .pdf
- **Capture video.** Record video, like 3D scatterplots, in various formats
- **To Notebook.** Sends the entire canvas to the Notebook.

- Sheets are shown in the lower left corner. To add a blank sheet select the plus icon. For sheet options, click on the arrowhead

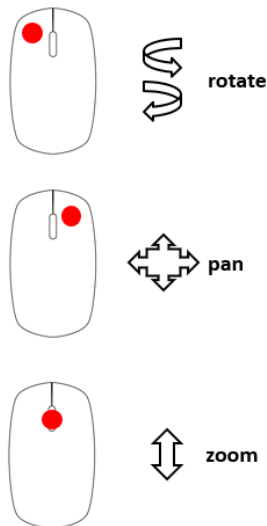


Customizing the Data Viewer Appearance

- Click on arrow icon to collapse or expand  
- Plots are separated by grippers. Mouse over a gripper to see options
 - Click and drag: resize the plot
- To reset the view of the plot, use the  control on the right of each plot
- To move a plot to a new position, use the handle at the top



- Mouse operation on a plot:



In-plot controls

- Plot tools are located in the upper right corner and appear upon a mouseover



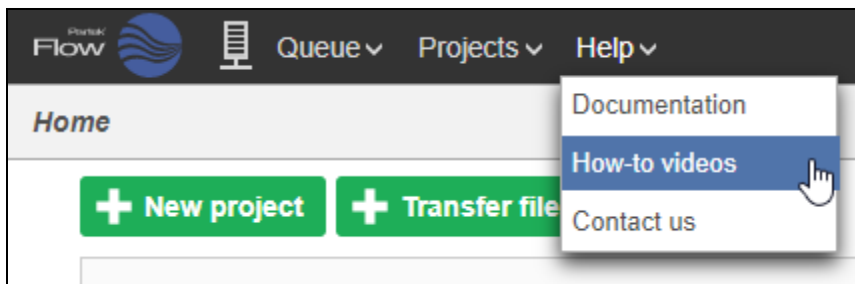
- Horizontal series
 - *Duplicate plot.* Makes an exact replica
 - *Save Image.* Saves the selected plot only as an image to the local computer. Supported formats are .png and .svg
 - *Send to Notebook Page.* Sends the selected plot to the Notebook
 - *Fullscreen plot/Exit Fullscreen mode.* Plot is expanded to fill the canvas
 - *Remove plot.* Removes the selected plot from the canvas. *Data Viewer* session remains open
- Vertical series
 - *Export data.* Download a .txt file with data use to draw a scatter plot (2D & 3D scatter plots only)
 - *Pointer mode.* Click to select data points
 - *Lasso mode.* Draws a lasso to select data points
 - *Invert Selection.* Unselects the currently selected cells, and selects the others
 - *Reset View.* Resets plot rotation and zoom
 - *Toggle Legend.* Turns legend on and off
 - *Toggle Axes Autoscale.* Scales the axes with respect to the visible data points (e.g. after some data points have been filtered out)
 - *Transpose.* Switch the axes



Getting Help

Self-learning

- Partek Flow documentation
<https://documentation.partek.com/display/FLOWDOC/Partek+Flow+Documentation>
- Step by step tutorials + practice data sets
<https://documentation.partek.com/display/FLOWDOC/Tutorials>
- Recorded webinars <https://www.partek.com/webinars/>
- Live training event recordings
<https://documentation.partek.com/display/FLOWDOC/Live+Training+Event+Recordings>
- Partek blog page <https://www.partek.com/blog/>
- Tips and tricks on Partek Flow are regularly tweeted
https://twitter.com/Partek_Inc
- *How-to videos* are accessible from the **Help** menu



Technical Support

- Open a support ticket at partek.com/support