

Frederick National Laboratory for Cancer Research



Applications of Single Cell Sequencing: CITE-Seq

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What is CITE-Seq?

- **“Cellular Indexing of Transcriptomes and Epitopes by Sequencing”**
- **Short definition: Including surface protein antibodies with single cell sequencing**
- **Purpose: Show that the antibody cell sorting often used prior to single cell sequencing is replicated after sequencing**
- **Need came up because the single cell transcript data does not always resemble the expected surface protein profile**

nature|methods

Brief Communication | Published: 31 July 2017

Simultaneous epitope and transcriptome measurement in single cells

Marlon Stoeckius , Christoph Hafemeister, William Stephenson, Brian Houck-Loomis, Pratip K Chattopadhyay, Harold Swerdlow, Rahul Satija & Peter Smibert

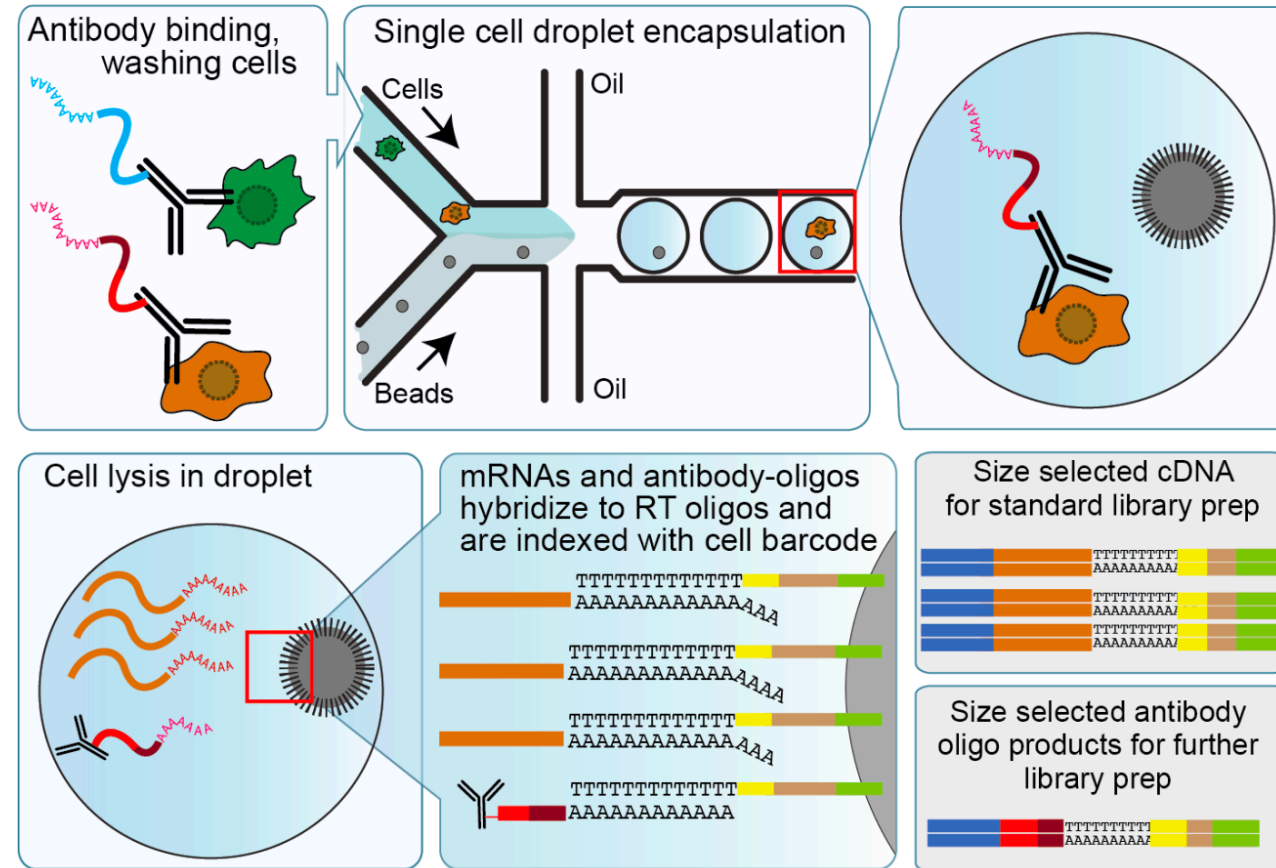
Nature Methods **14**, 865–868 (2017) | [Download Citation](#) ↓

Methodology

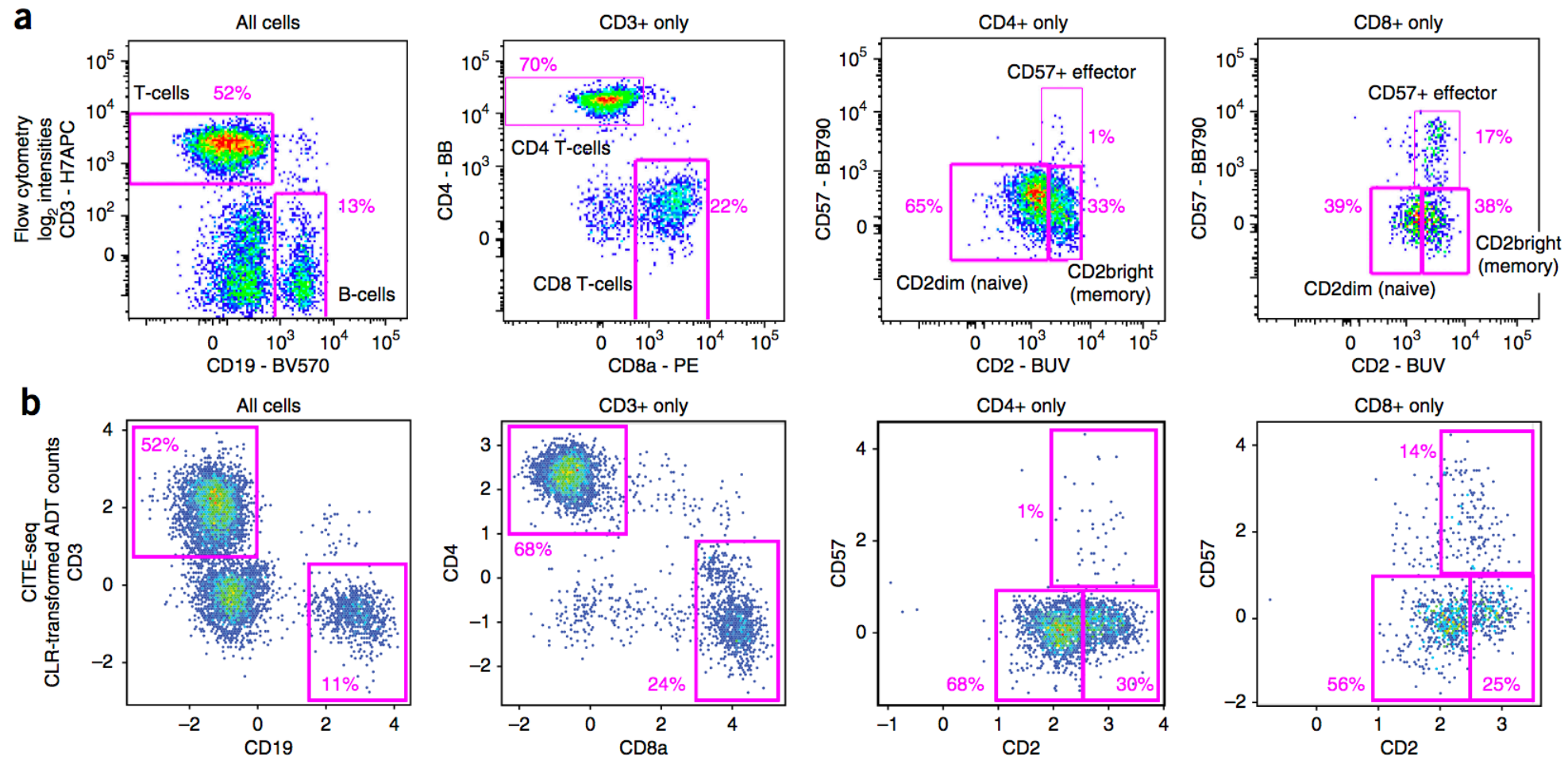
- **Specific antibodies with sequence tags are bound to cells prior to single cell encapsulation**



- **Each droplet contains both the cell and the bound antibody tag**
- **mRNA and antibody oligos hybridize to RT oligos and are indexed with cell barcode**
- **Standard scRNASeq is performed for mRNAs**
- **Antibody tags are isolated by size selection**

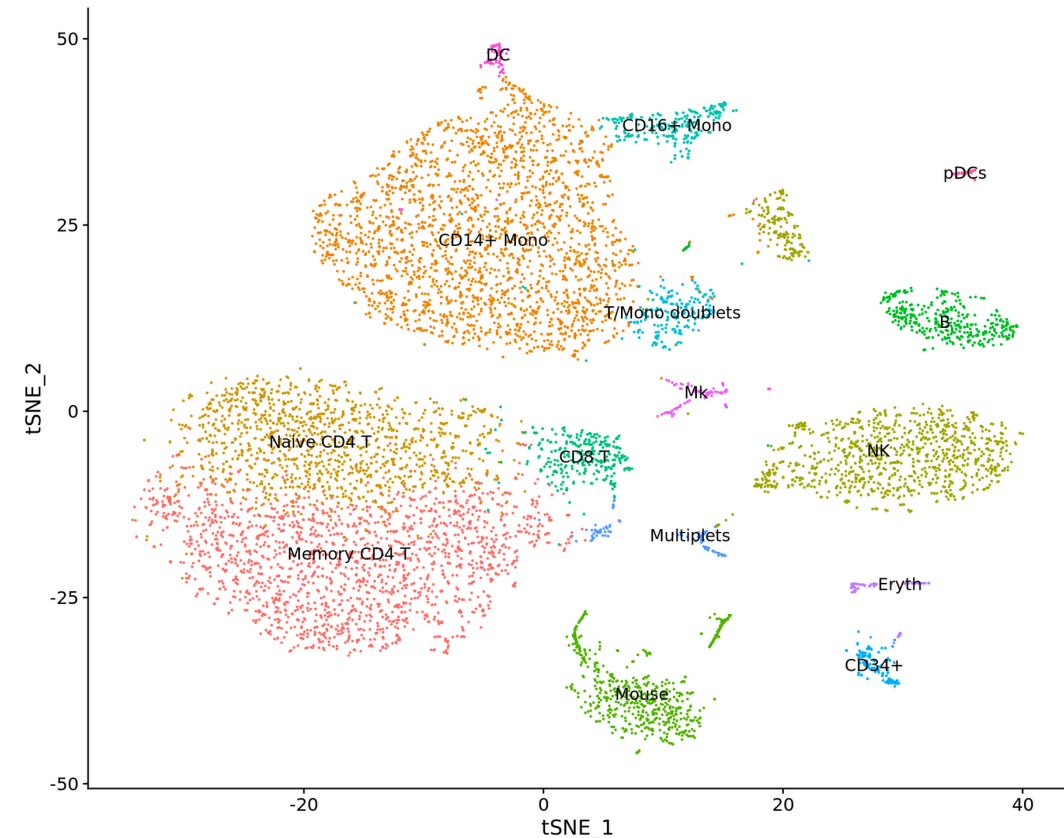


CITE-Seq is comparable to initial flow cytometry



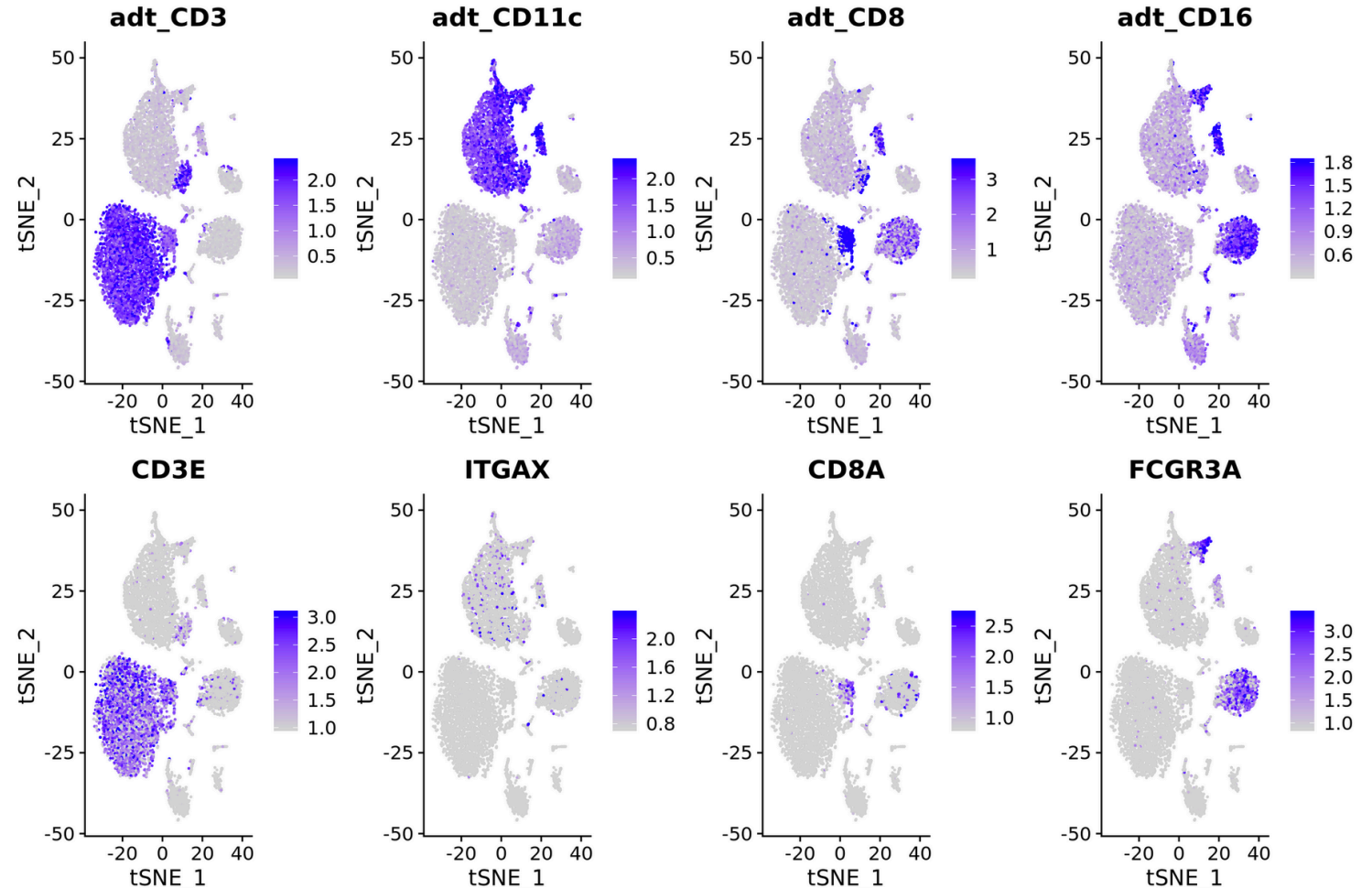
Antibody-based identification can be projected onto RNA-Seq based clustering

- Initial reference image is processed using “standard” scRNA-Seq protocol
- Labels for this image were assigned by the Satija group based on prior knowledge of the dataset
- CITE-Seq protein markers for this dataset include:
 - CD3, CD4, CD11c, CD14, CD16, CD19 CD34, CD45RA, and CD56

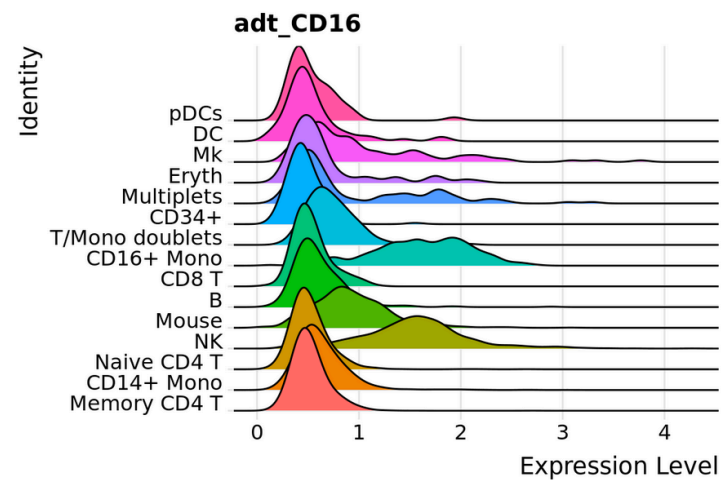
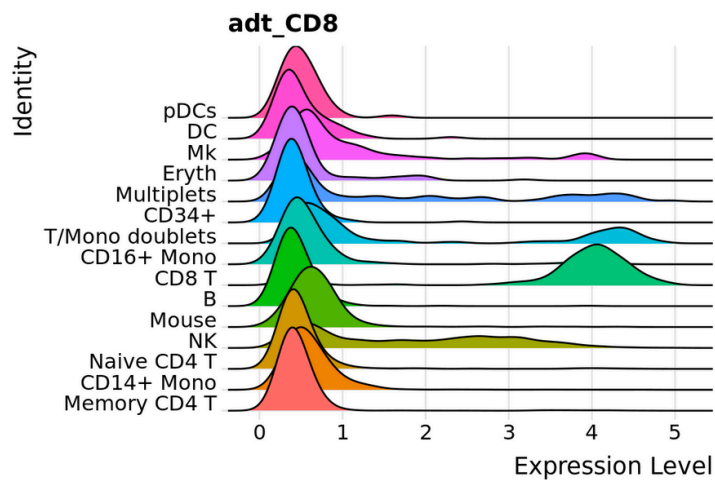
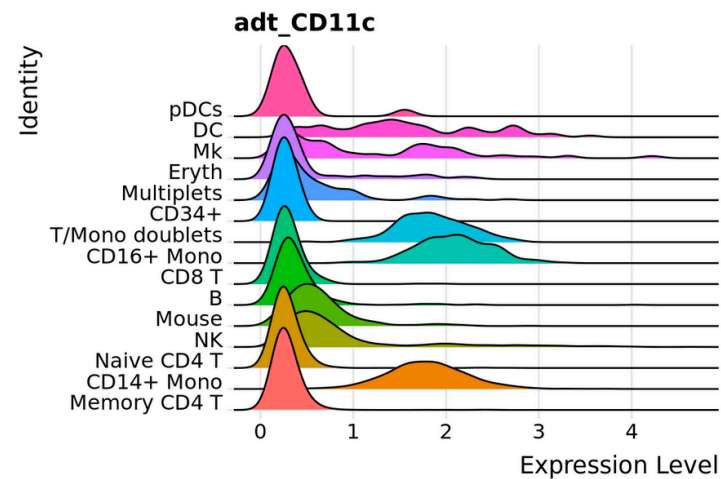
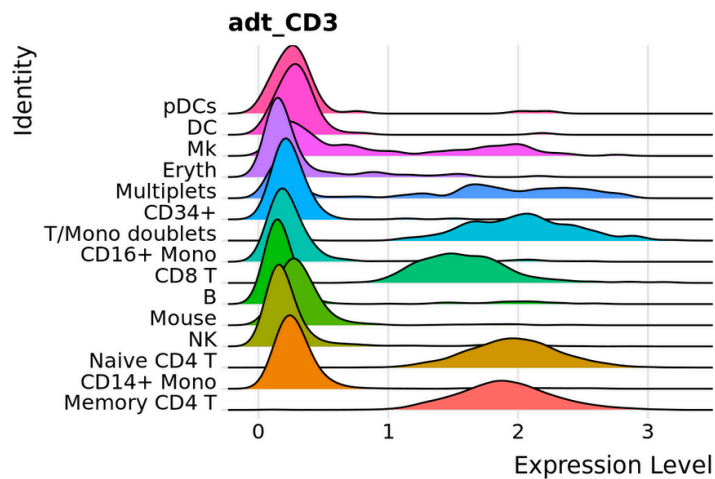


Antibody-based identification can be projected onto RNA-Seq based clustering

- **Top: Protein markers**
- **Bottom: Associated gene IDs**
- **Using combined biological knowledge and protein expression allows for the confirmation of individual cell types in scRNA-Seq**
- **Cell surface markers can confirm the RNA-Seq-based cell typing**

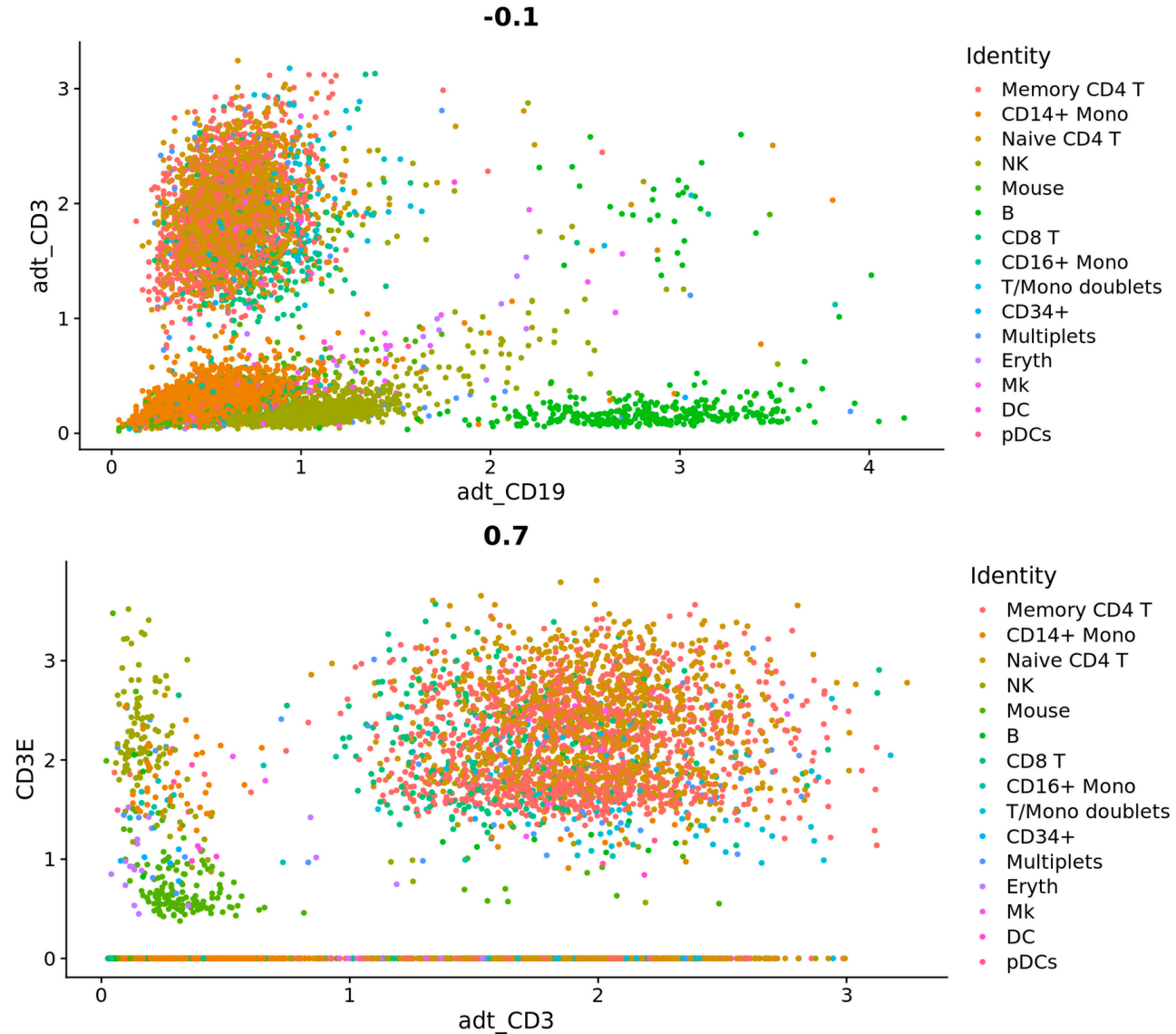


Visualizing protein surface marker binding can confirm cell identities



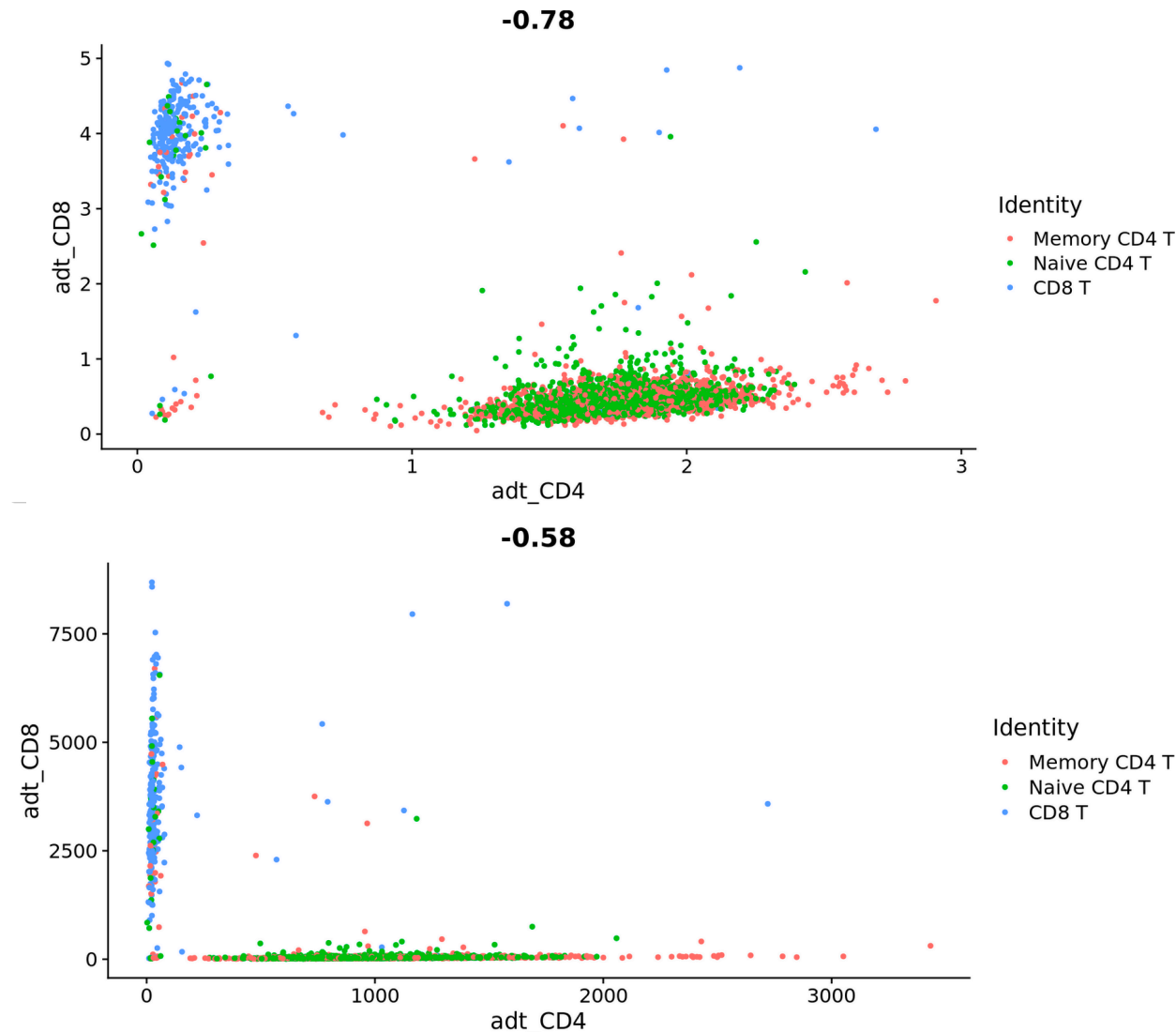
Scatter plots and correlation analysis can be used to show “protein vs. protein” and “protein vs. RNA”

- Interest in surface protein expression can show positive, negative, or null correlation
- The previous image for surface protein vs. transcript expression can be indicated with scatter plots as well
 - Note that the gene expression has more zero values than the protein marker



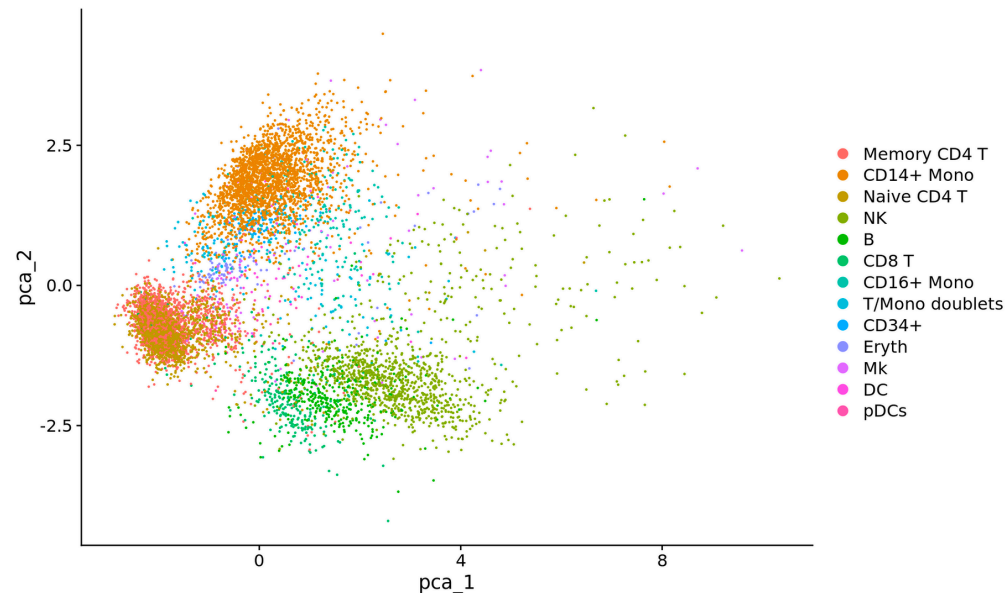
Protein markers can improve cell typing

- **CD4 and CD8 markers are projected for T-cells**
- **Some cells identified transcriptomically as CD4+ T-cells are being shown in the CD8+ protein marker population**
- **Biologically, this is not unexpected, because naïve CD4+ T-cells and CD8+ T-cells have similar transcriptomes**
- **This could have larger effects on downstream analysis without CITE-Seq reinforcement**

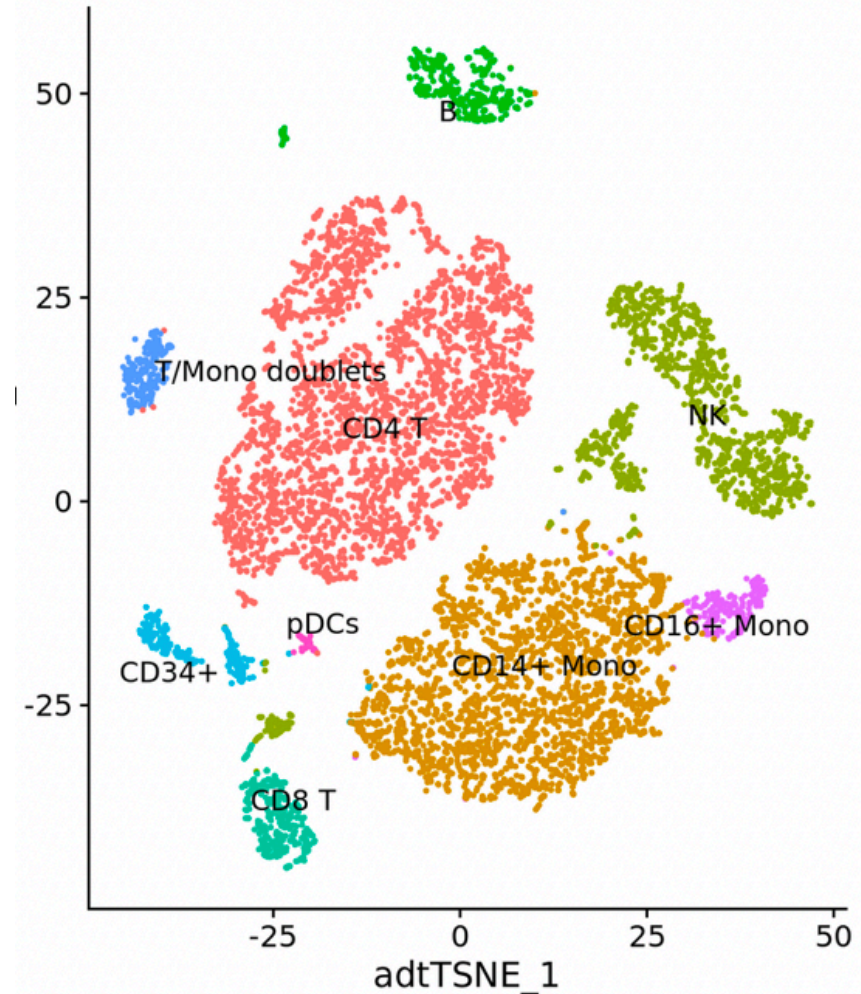


Clustering can be performed directly on the protein expression profile

- Many of the analyses performed based on scRNASeq (transcriptome) can be performed based on CITESeq
- The more markers that are used, the better the profiling/clustering that can be extracted

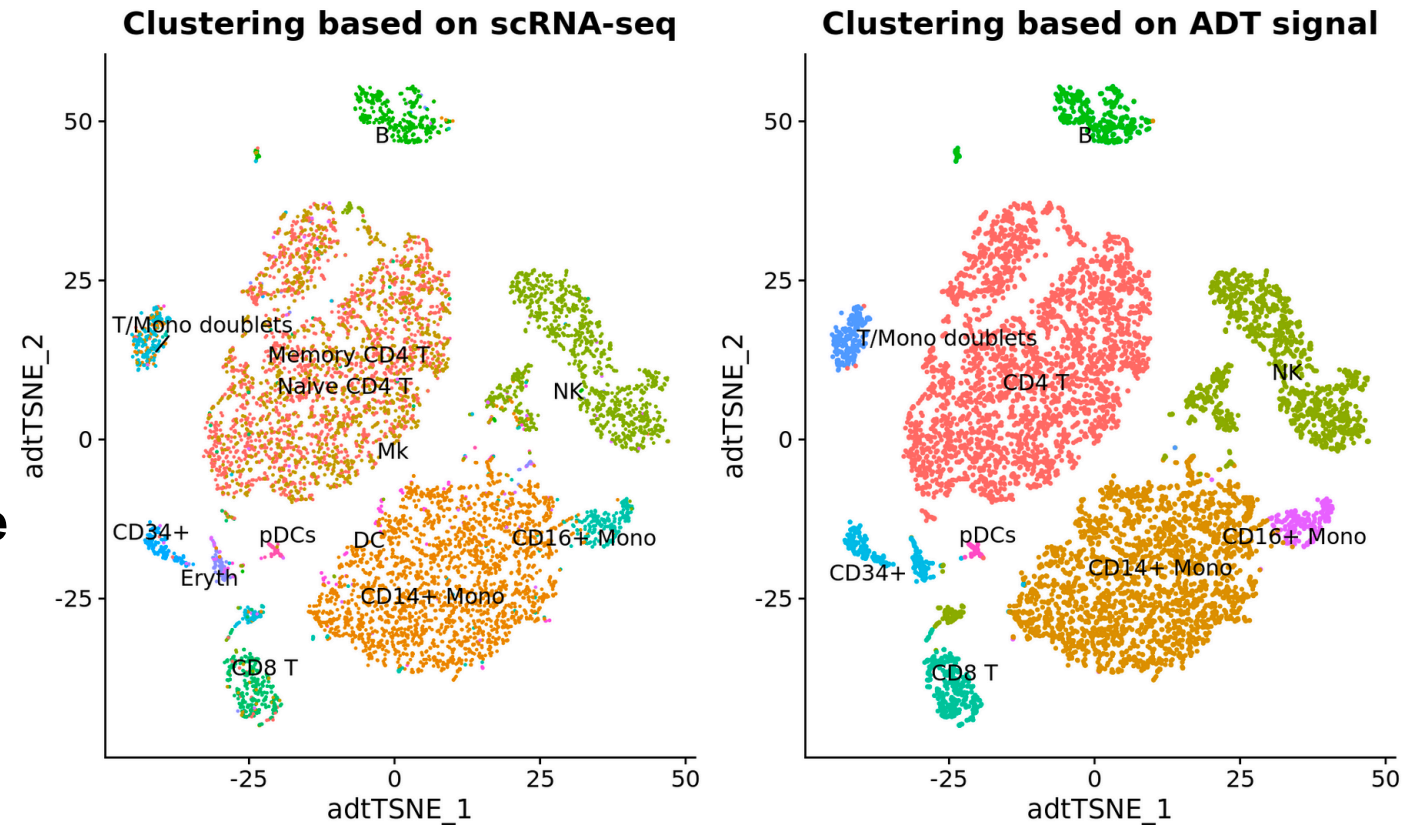


Clustering based on ADT signal



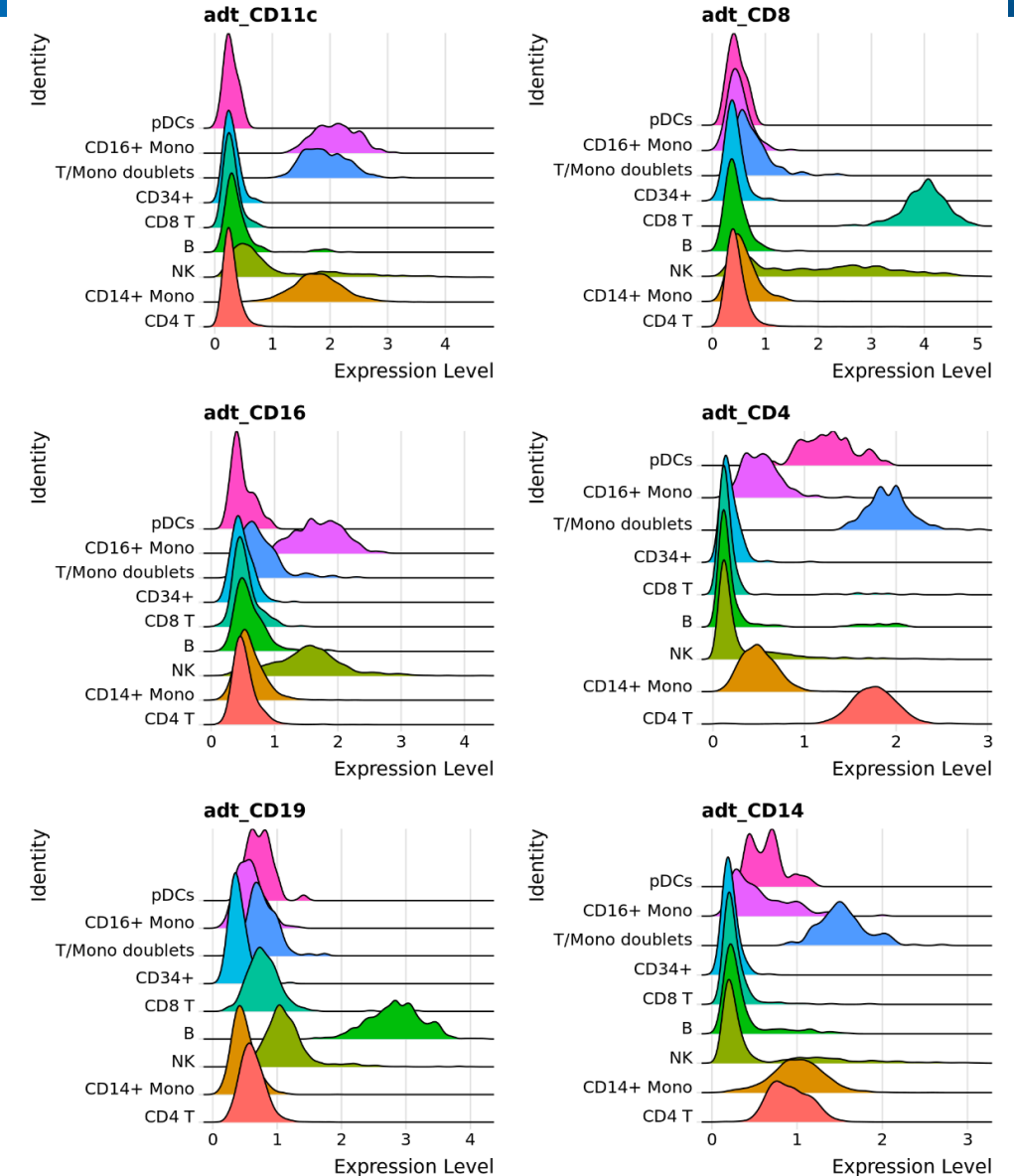
Clustering between scRNA-Seq and CITE-Seq/protein markers generate similar results

- Cluster positions in this image are based on the protein signal
- The labels on the left are based on the scRNA signal, the labels on the right based on protein signal
- Advantage: The CD4+ and CD8+ T-cells are more clearly defined on the protein signal
- Disadvantage: Some cell types with poor/absent markers (e.g. erythrocytes) are lost



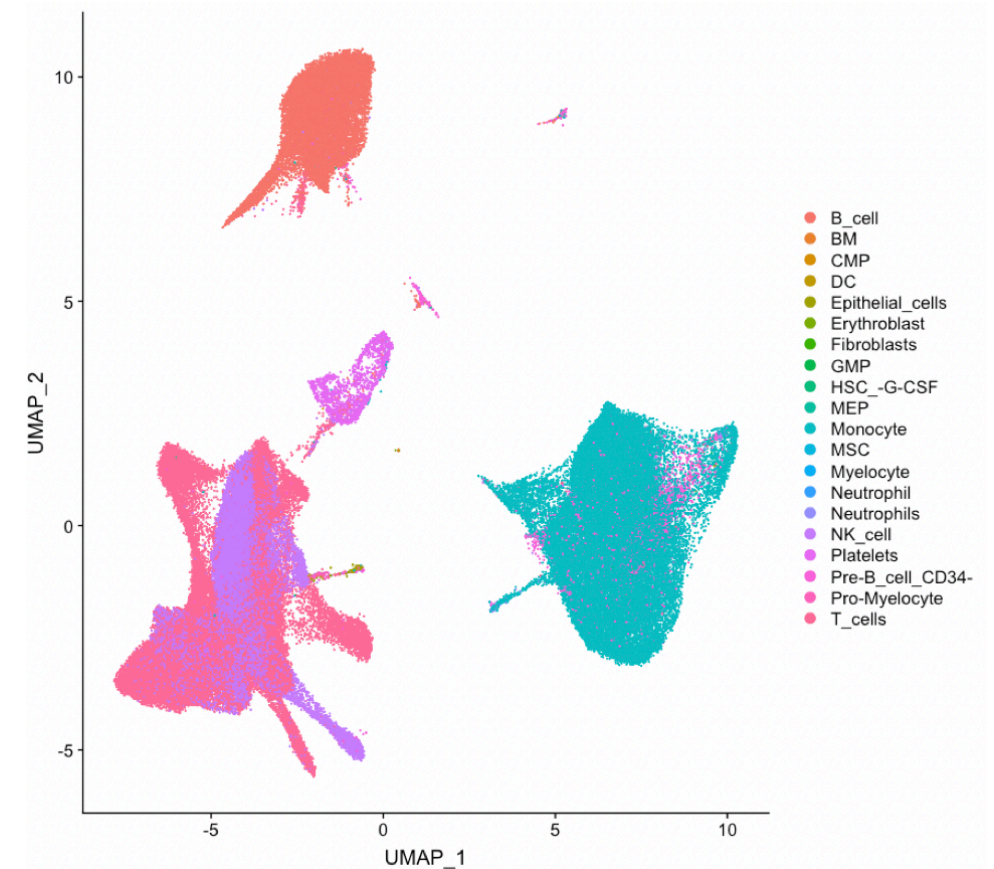
Surface marker profiles are representative of the cell types

- The ridge plot can show the relative expression of the proteins for each cluster ID
- Based on these results, it would be similar to using FACS in determining cell typing

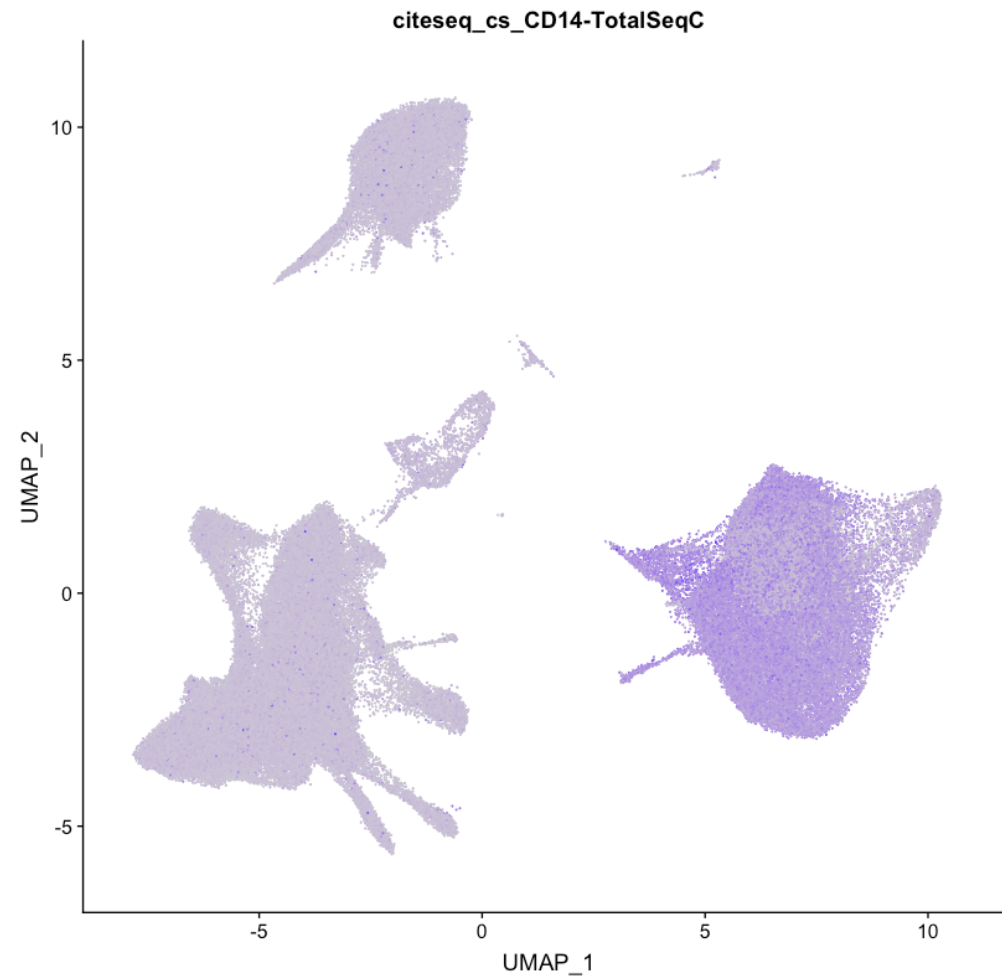
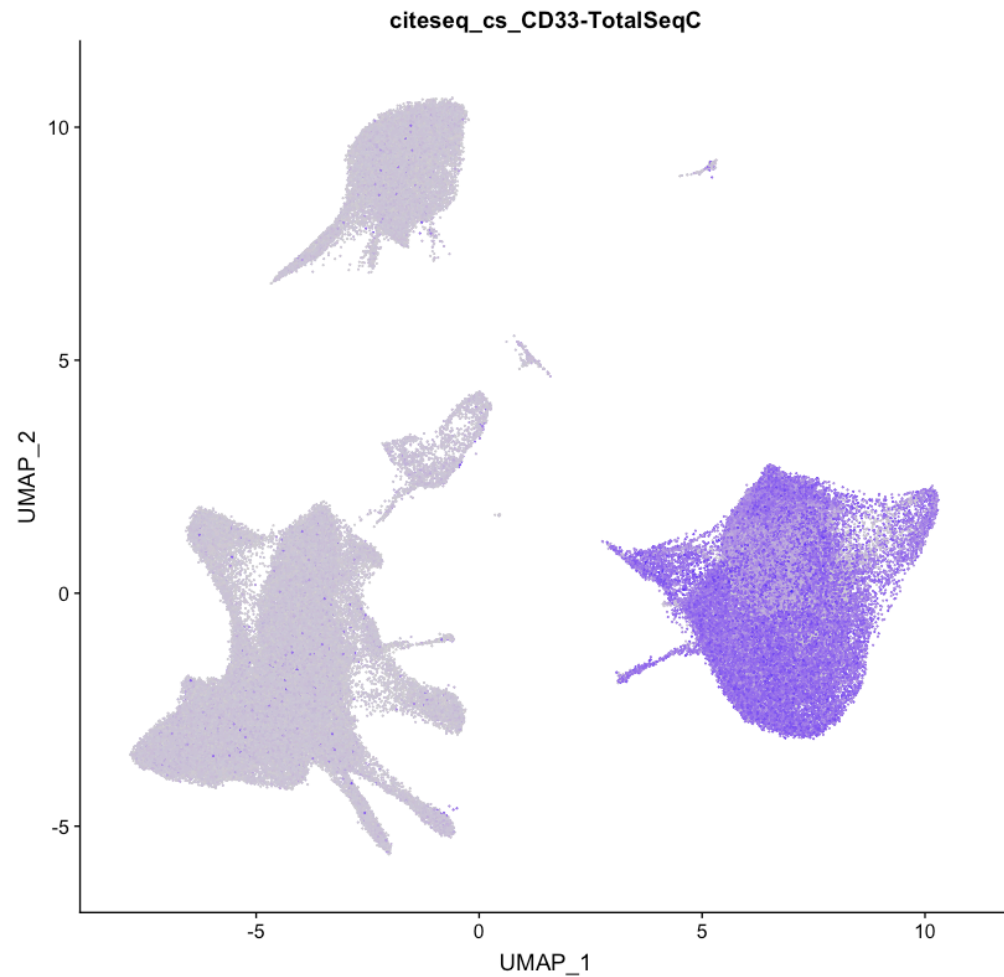


“Live” application of CITE-Seq

- **Short background: Human PBMCs from 11 different samples**
- **Interest in CD33 and CD14 as potential surface markers**
- **The initial transcript based cell-clustering and cell typing with SingleR showed the majority of cells as:**
 - B-cells (top)
 - T-cells and NK-cells (bottom left)
 - Monocytes (bottom right)

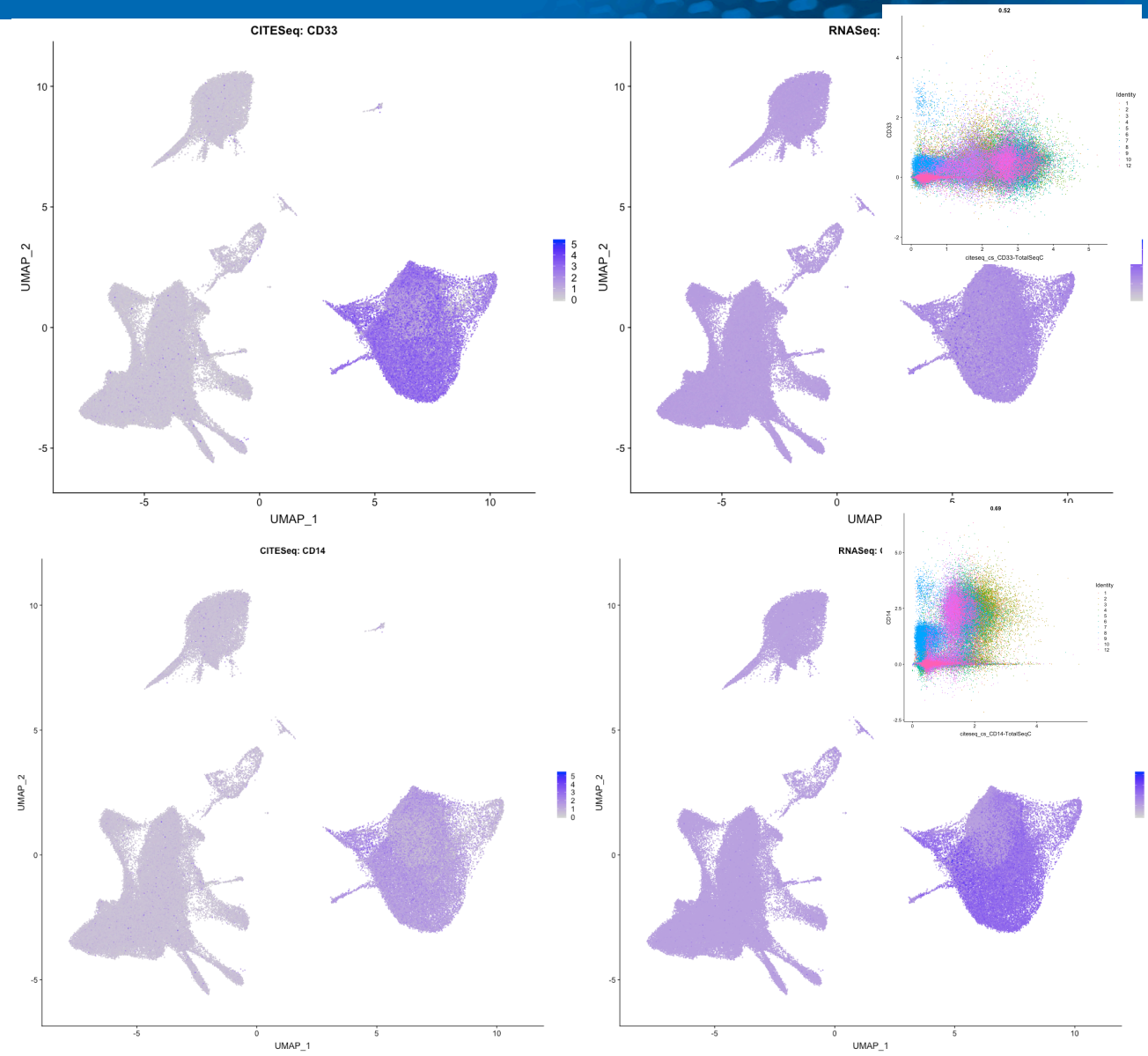


CD33 and CD14 are strong correlative markers for monocytes



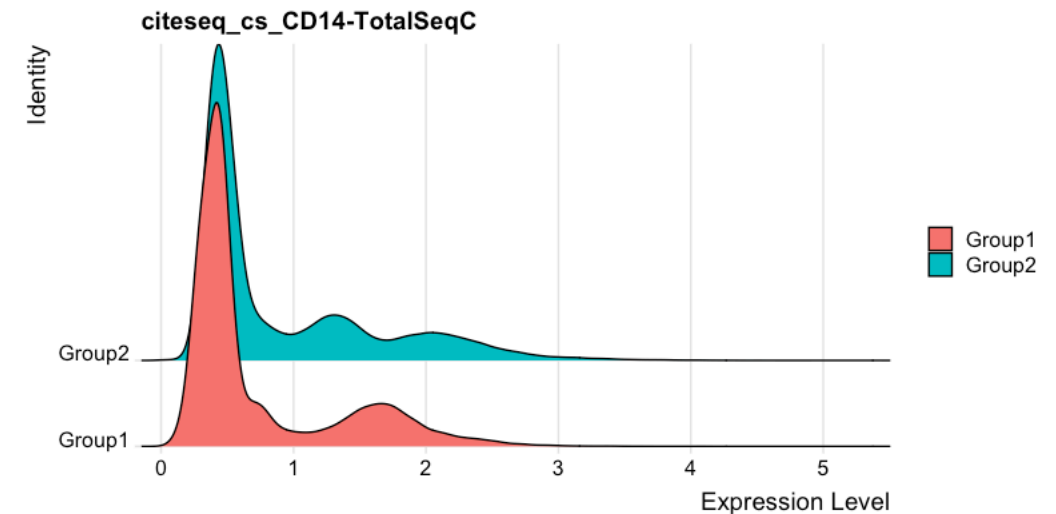
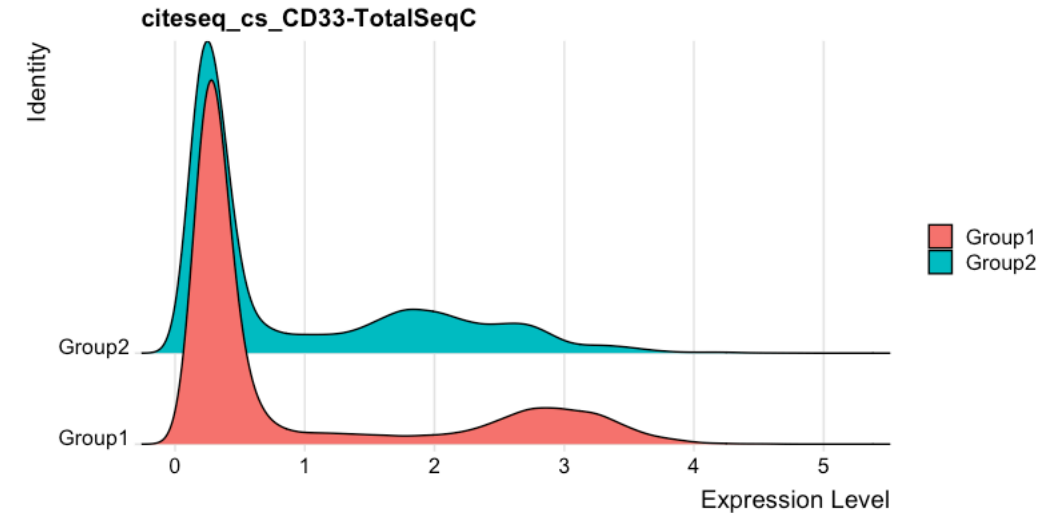
CITESeq indicates a discrepancy between transcription and surface protein accessibility

- Protein profiles (left) showed more noticeably “antibody responsive” markers in monocytes
- The difference is less pronounced at the transcription level (right), especially for CD33
- This is even more apparent when observing the scatter plots (insets)



CITESeq for identifying sample groups

- **Samples were split into two categories.**
- **Group 1 showed more definite CD33 surface protein**
- **CD14 was confirmed as a less readily usable marker in defining the two groups**



Take home message

- **CITE-Seq is an excellent tool for integrating traditional antibody cell sorting with scRNA-Seq**
- **This can help synchronize the FACS results with scRNA-Seq**
- **Word of warning: Understand the underlying biology**
- **Per the Hitchhiker's Guide to the Galaxy: Don't Panic**

References

- Seurat Guided Analyses. Satija R, et al. <https://satijalab.org/seurat/vignettes.html>
- Stoeckius M, Hafemeister C, Stephenson W, Houck-Loomis B, Swerdlow H, Satija R, Smibert P. 2017. [Simultaneous measurement of epitopes and transcriptomes in single cells.](#) *Nature Methods*.
- Butler A, Hoffman P, Smibert P, Papalexi E, Satija R. 2018. [Integrating single-cell transcriptomic data across different conditions, technologies, and species.](#) *Nature Biotechnology*.
- Stuart T, Satija R. 2019. [Integrative single-cell analysis.](#) *Nature Reviews Genetics*.
- Stuart T*, Butler A*, Hoffman P, Hafemeister C, Papalexi E, Mauck WM, Hao Y, Stoeckius M, Smibert P, Satija R. 2019. [Comprehensive Integration of Single-Cell Data.](#) *Cell*.