

# Methylation Array Analysis

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# Partek® Genomic Suite™ Main Dialog

**Analytical spreadsheet:** Central repository of data

- No limitation on number of rows or columns
- Rows represent observations of interest (experiments, samples, chips)
- Columns represent measures of the observations (variables, features, genes,)

**Menu bar:** Execute commands from a graphical user interface

- When spreadsheet is empty, most of the menu items are not displayed

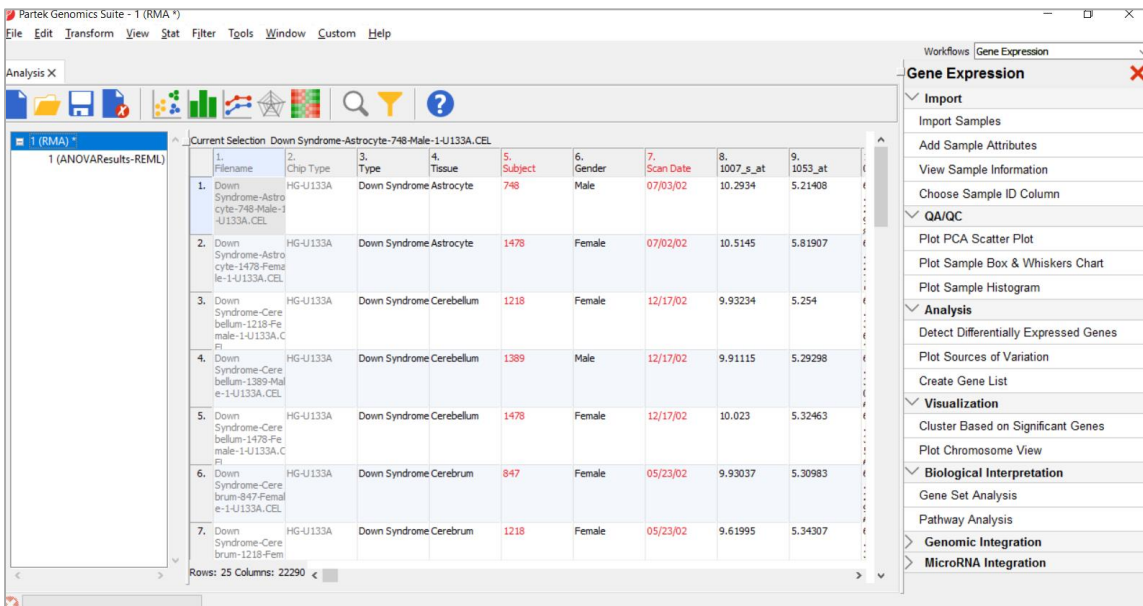
**Tool bar:** Accelerator buttons allow quick access to commonly used commands

**Spreadsheet hierarchy:** Open multiple datasets and see the hierarchy

- Original spreadsheet: parent
- Result spreadsheet: child

**Active spreadsheet:** The active spreadsheet is shown highlighted in blue, and the spreadsheet name and associated file name are shown at the top of the dialog

**Workflow:** Used to guide you through a typical analysis of a specific assay



**Notes:**

# Tutorial Data Set

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- The data set for the exercise is based on Gene Expression Omnibus GSE38240
- Download data from:
  - [https://customer.partek.com/Methylation\\_training.zip](https://customer.partek.com/Methylation_training.zip)
- Aryee *et al.* DNA methylation alterations exhibit intra-individual stability and inter-individual heterogeneity in prostate cancer metastases. *Sci Transl Med* 2013 Jan 23;5(169):169ra10.
- Prostate samples from
  - Normal individuals
  - Those diagnosed with prostate cancer
- Profiled using Illumina HumanMethylation450 BeadChip
- The goal of the exercise is to come up with a list of genes that show evidence of hyper- or hypo-methylation in tumor comparing to normal in promoter regions

**Notes:** \_\_\_\_\_

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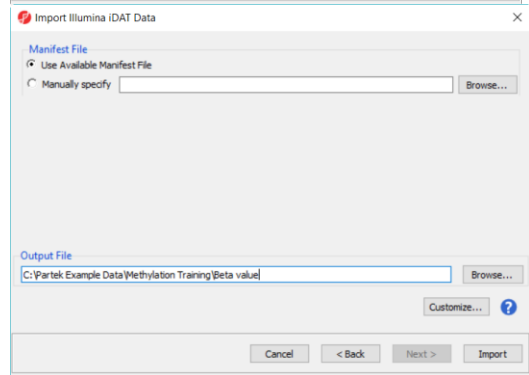
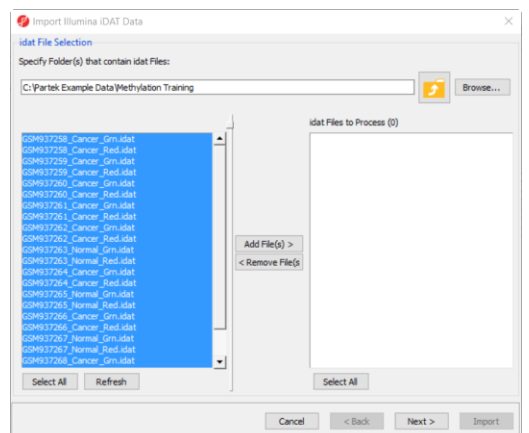
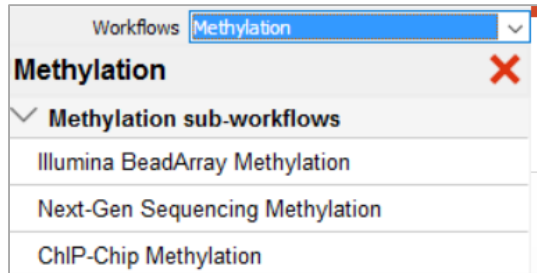
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# Importing .iDAT Files

- Set the *Workflows* selector to **Methylation**
- Select **Illumina BeadArray Methylation**
- Select **Import Illumina Methylation Data**
- In the pop-up window, select **Import human methylation 450/850 .idat files**
- Browse to the folder, add the files to the right panel by clicking **Add Files**, there are 24 files to process
- Click **Next>**
- **Use Available Manifest File** option, name the output file as *GSE38240 data*, click **Import**
- The needed library file will be automatically downloaded
- The default is using functional normalization to generate  $\beta$ -values, which correspond to the percentage of methylation at each site
  - Ratio of methylated probe intensity over the overall intensity at each site.
- Each row of the spreadsheet corresponds to a single sample with the methylation probes on columns



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# Convert Beta value to M Value

- Click **Convert Beta Value to M Values**
  - $M\text{-value} = \log_2(\beta / (1 - \beta))$
- The spreadsheet is overwritten with M value, click **Save**
- M value interpretation:
  - a M-value close to 0 indicates a similar intensity between the methylated and unmethylated probes, which means the CpG site is about half-methylated. Positive M-values mean that more molecules are methylated than unmethylated, while negative M-values mean that more molecules are unmethylated than methylated.
  - the M-value is more statistically valid for the differential analysis of methylation levels.

Current Selection GSM937258_Cancer								
	1. Sample ID	2. cg00050873	3. cg00212031	4. cg00213748	5. cg00214611	6. cg00455876	7. cg01707559	8. cg02011394
1.	GSM937258_Cancer	-0.277581	-2.71171	-3.0262	-4.26836	1.83445	-2.35141	0.720438
2.	GSM937259_Ca	0.126901	3.22854	-0.355054	-3.98109	-0.439771	-2.86905	0.72438
3.	GSM937260_Ca	0.0346121	-4.97955	2.03609	-5.10928	1.87764	-4.22799	5.16259
4.	GSM937261_Ca	0.0388291	2.91757	2.44367	-4.16336	-0.363247	-2.79682	0.859743
5.	GSM937262_Ca	1.12966	1.54597	1.01182	-5.03341	0.380025	-4.2639	2.56671
6.	GSM937263_No	2.55891	-4.91436	2.04838	-4.83353	1.1223	-4.4994	5.95356
7.	GSM937264_Ca	0.118948	3.11273	-3.3809	-4.74306	-1.33189	-3.33236	1.37507
8.	GSM937265_No	2.51931	-5.28035	2.43734	-4.90895	1.82947	-4.51953	5.65953
9.	GSM937266_Ca	3.96307	-0.497761	-0.311449	-0.575516	-0.135092	-0.816626	5.49651
10.	GSM937267_No	2.25543	-5.13274	2.36759	-5.1285	1.41825	-3.58654	5.83423
11.	GSM937268_Ca	2.59777	-0.170689	-0.531692	-0.205582	0.71798	-1.00491	5.38137
12.	GSM937269_No	3.09111	-4.81732	2.02696	-4.64226	1.8368	-4.70668	6.09491

Rows: 12 Columns: 485513

**Notes:** \_\_\_\_\_

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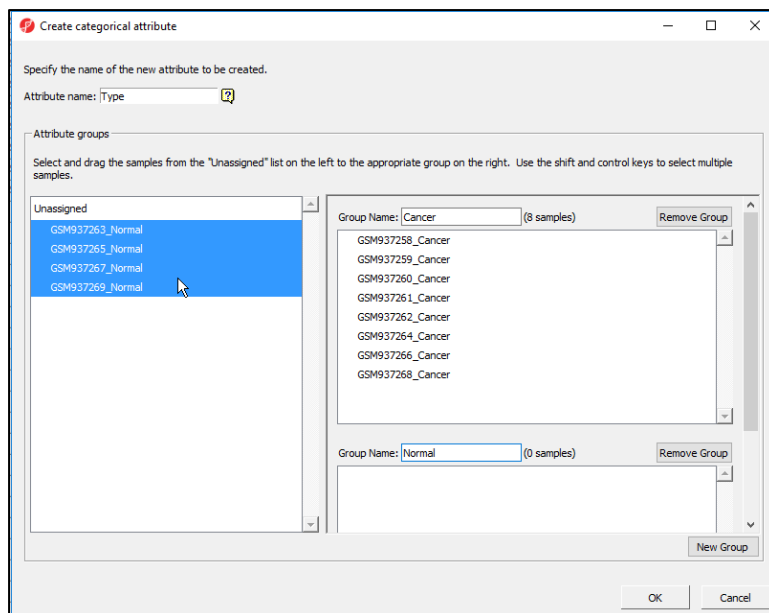
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# Annotating Samples

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- Click **Add sample attributes > Add a categorical attribute > OK**
- In the dialog, set the *Attribute name* to **Type**, change the label *Group 1* to **Cancer** and *Group 2* to **Normal**
- **Ctrl-select** the samples labeled *Cancer* and **drag and drop** to the *Cancer* group
- **Drag and drop** the remaining samples to the *Normal* group. Click **OK**.
- When prompted to *Add another attribute*, click **No**
- Save spreadsheet with the new sample attribute, click **Yes**



**Notes:** \_\_\_\_\_

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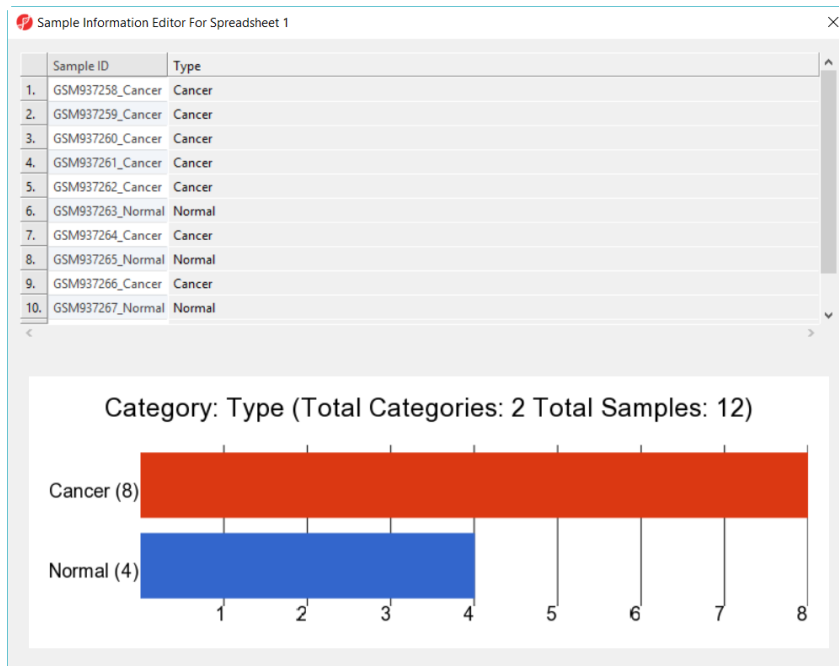
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# View Sample Information

- Click **View Sample Information** on the workflow
- There are 8 Cancer samples, 4 Normal samples
- Choose Sample ID column: use the default, it has to be unique ID for each sample



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# Exploratory Analysis

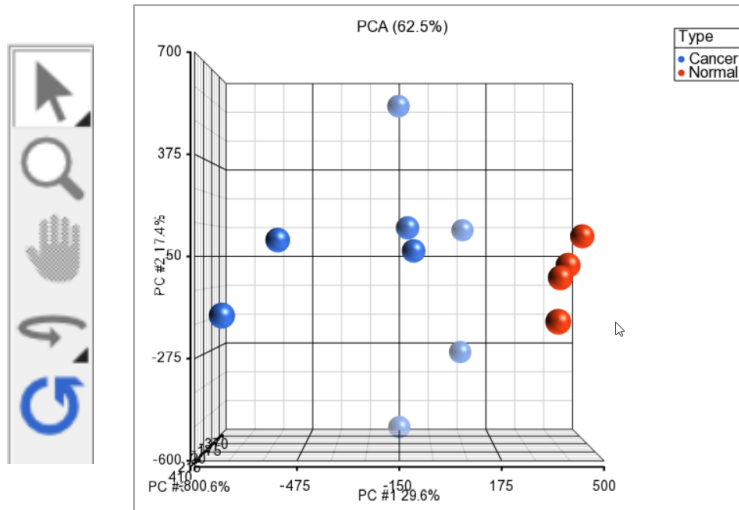
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PCA scatter plot is one way to identify clustering patterns and outliers

- Go to the QA/QC setion of the workflow > **Plot PCA Scatter Plot**

## Notes

- Each point in the scatter plot corresponds to a specific row in the spreadsheet
  - Points that are close together in the plot are similar in the original high-dimensional space
  - Points that are far apart in the plot are dissimilar
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- Click on **Plot Properties** to configure color
  - Click on **Ellipsoid** to put the ellipsoid on each group
  - Select mode:
    - left click to select; scroll mouse wheel to zoom; drag mouse wheel to rotate
    - right click after select a point to filter/clear filter

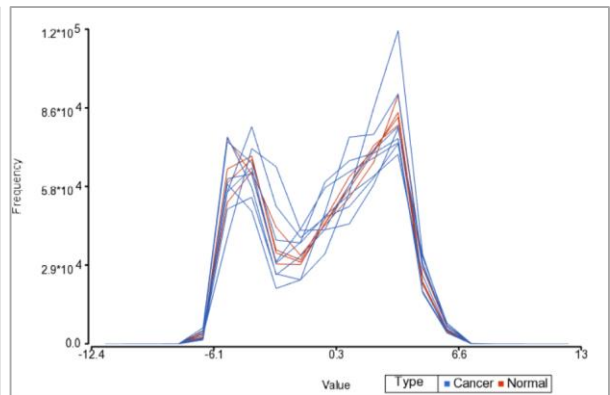
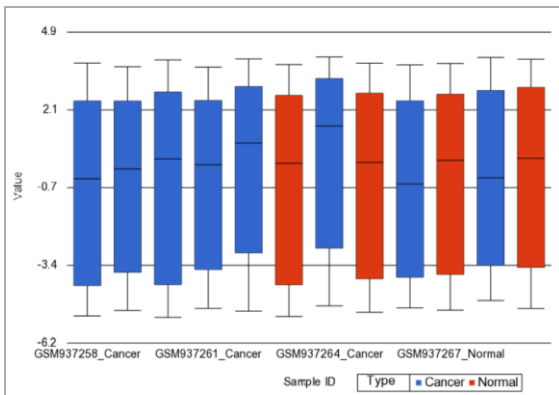


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# QA/QC – Histogram and Box plot

- Select **Plot Sample Box & Whiskers Chart**
  - Each box is a sample
  - Line inside the box is the median (2<sup>nd</sup> quartile)
  - Box represent the first and third quartiles
  - Whiskers represent 10<sup>th</sup> percentile and 90<sup>th</sup> percentile by default, can be configured
- Select **Plot sample histogram**
  - Each line is a sample
  - X-axis is the range of the values
  - Default 20 bins on X-axis, can be configured from **Plot Properties**



**Notes:**

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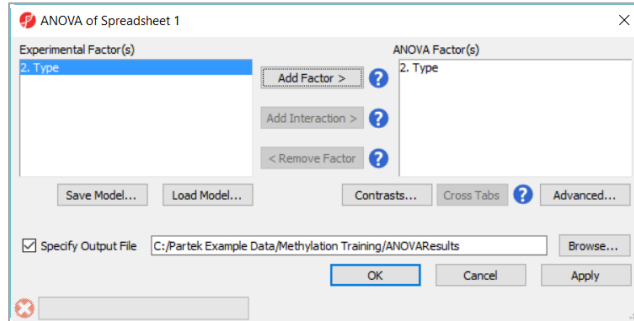
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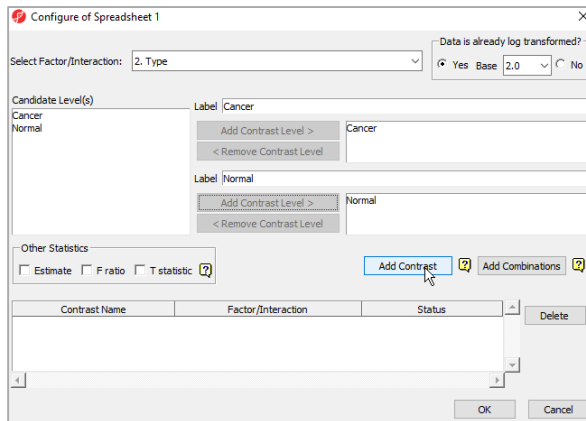
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# Detecting Differentially Methylated Loci

- Go to **Analysis > Detect differential methylation**
- Select **2. Type** under *Experimental factor* and click the **Add Factor>** button
- Click the **Contrast** button



- Choose **Yes** for *Data is already log transformed*
- Use **Add Contrast Level>** to move **Cancer** to *Group 1* and **Normal** to *Group 2*
- Select **Add Contrast** and then **OK**. In the ANOVA dialog also click **OK**.

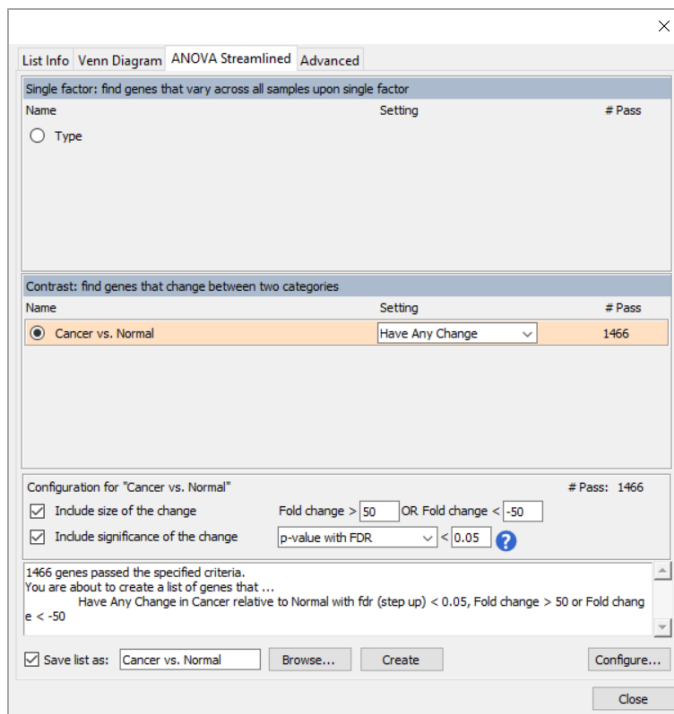


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# Creating marker list

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- Click **Create marker list** from the workflow
- Select the **Cancer vs Normal** radio button
- Set the size of the *Fold Change* filter to  $>50$  and  $<-50$ 
  - This selects markers that are either hyper- or hypo-methylated in Cancer comparing to normal
- Set the significance threshold to *p-value with FDR*  $<0.05$
- Click the **Create** button to make a new list with these filtered markers



**Notes:** \_\_\_\_\_

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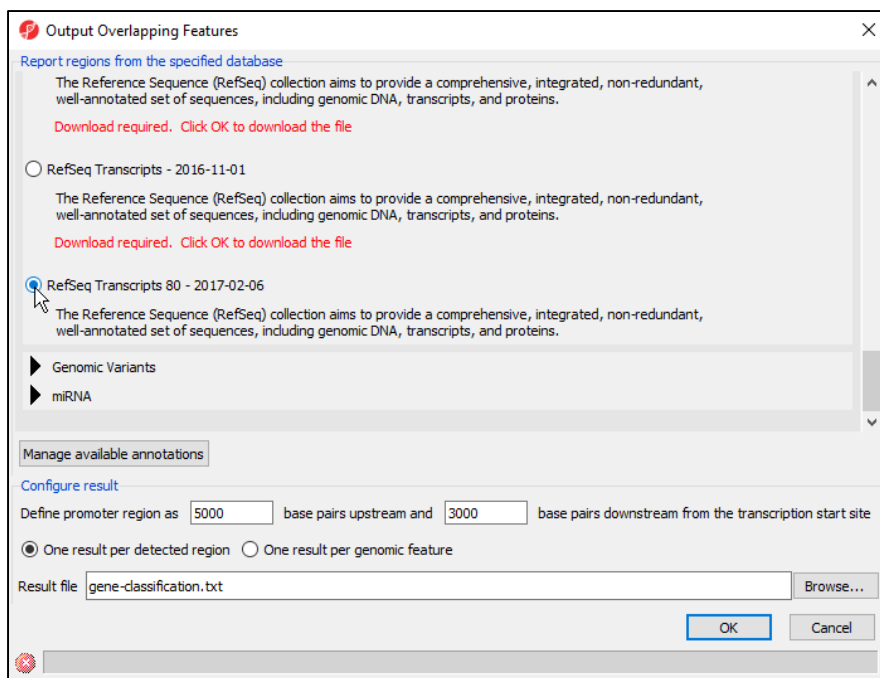
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# Classify regions by gene section

- With the *Cancer vs Normal* spreadsheet select **Classify regions by gene section** in the workflow
- Select the *RefSeq Transcripts 80 – 2017-02-06* radio button and click **OK**
- Using the default settings, the output spreadsheet contains each row is a probe overlap with a gene section
- One location can overlap with multiple transcripts



**Notes:**

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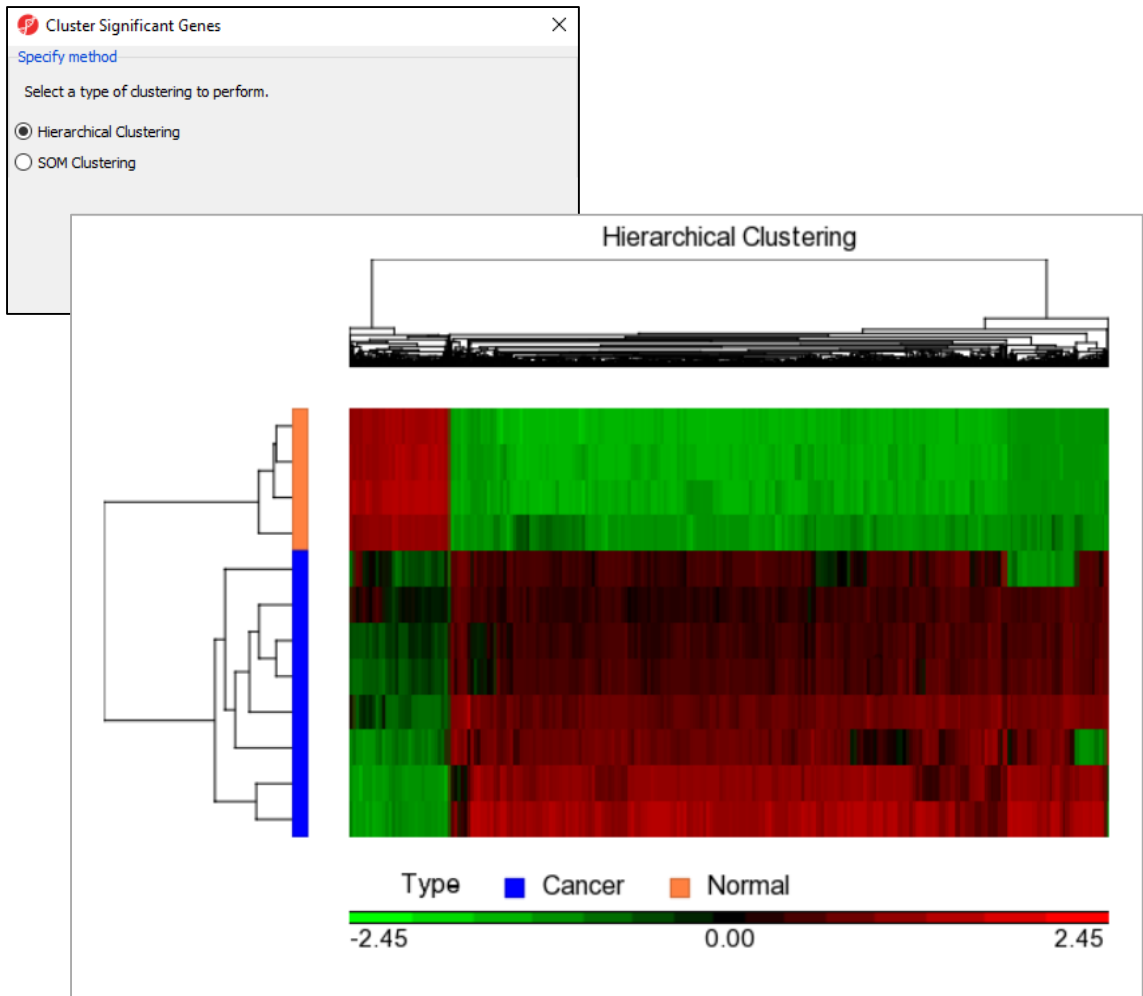
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# Cluster based on significant genes

- With the *Cancer vs Normal* spreadsheet select **Cluster based on significant genes** in the workflow
- Select **Hierarchical Clustering** and run the default settings



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# Hierarchical Clustering Configuration

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## Heatmap

- Click on the color square to change the heatmap color
- Change the orientation

## Dendrograms

- Change the width/height of the dendrogram
- Color dendrogram

## Rows

- Change the width of annotation
- Check show label
- Change color
- Add new annotation

## Columns

- Label with column header or gene symbol

**Save/Load:** save or load configuration settings

**Mode:** mouse over, select, zoom, and flip

**Notes:** \_\_\_\_\_


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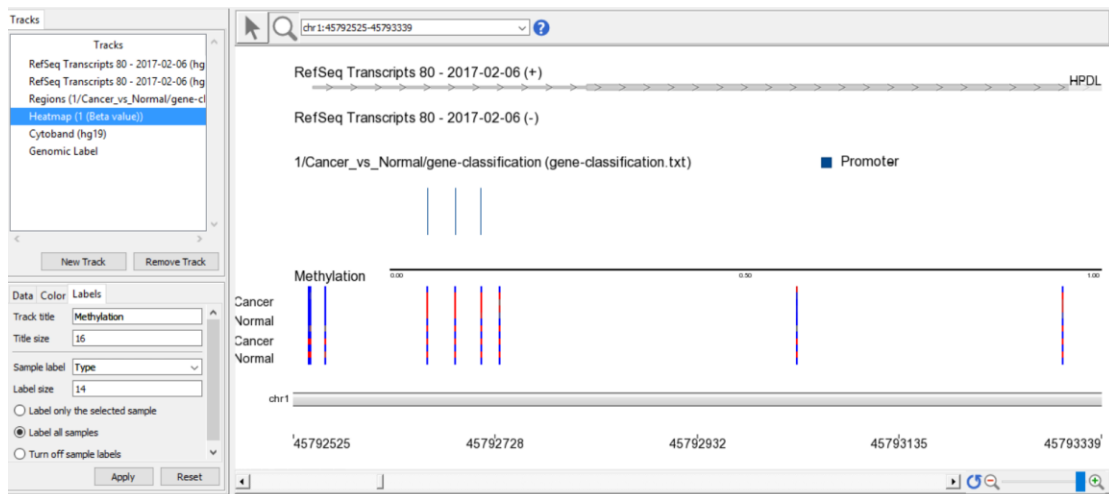
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# Viewing probes using chromosome view

- In the *gene-classification* spreadsheet, click the **interactive filter** button 
- The interactive filter gives a graphical representation of the values within a dataset and makes it easy to select values to filter
- Select **7. Gene Section** from the drop-down menu
- **Right click** on the rightmost bar, representing *Promoter*
- Choose Plot Chromosome View to visualize the result
- Select each track to change the configuration:
  - Remove the track of Cancer vs Normal list
  - Select region track of gene classification, change the Separate bars by to **None**
- In zoom mode, click and drag a region to zoom in



## Notes:

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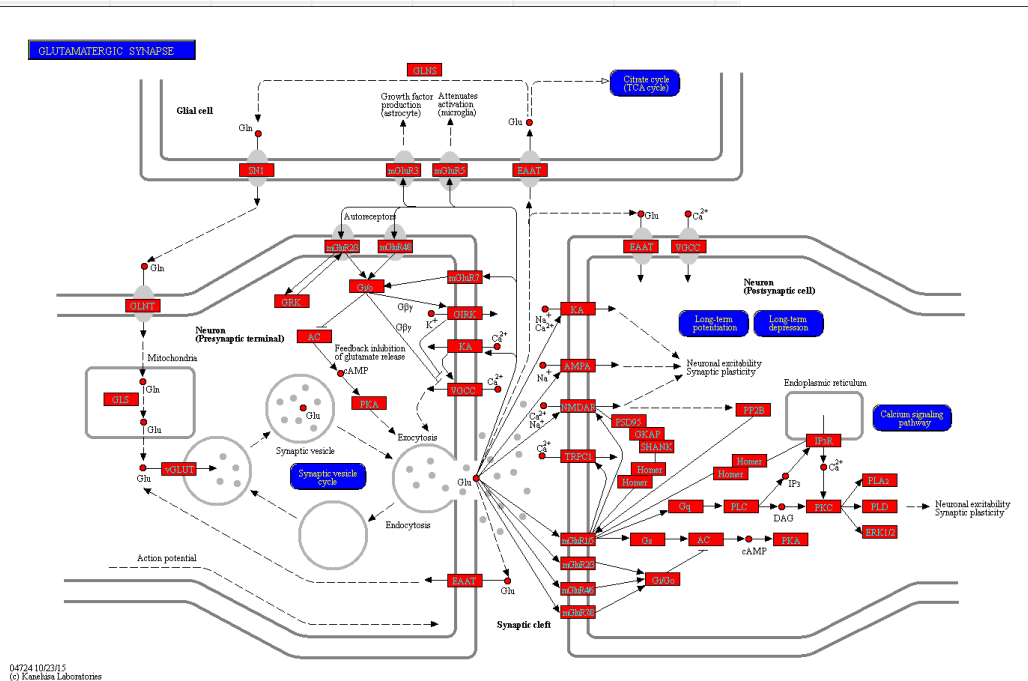
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# Pathway Enrichment Analysis

- With the *gene-classification* spreadsheet select **Pathway analysis** in the workflow.
- Select the **Pathway Enrichment** radio button
- The result spreadsheet contains each row representing a pathway with enrichment score

1.	Pathway Name	2. Database	3. Enrichment Score	4. Enrichment p-value	5. % genes in pathway that are present	6. # genes in list, in pathway	7. # genes not in list, in pathway	8. # genes in
1.	Glutamatergic synapse	kegg	7.30985	0.000668919	9.73451	11	102	197
2.	Long-term potentiation	kegg	7.0457	0.000871146	12.1212	8	58	200
3.	Retrograde endocannabinoid	kegg	6.88209	0.001026	9.90099	10	91	198
4.	N-Glycan biosynthesis	kegg	5.71402	0.00329937	12.5	6	42	202
5.	Neuroactive ligand-receptor	kegg	5.41087	0.00446776	6.20438	17	257	191
6.	Nicotine addiction	kegg	4.93261	0.00720767	12.5	5	35	203
7.	Gastric acid secretion	kegg	4.90288	0.00742518	9.45946	7	67	201

8. Calcium signa
9. Long-term de
10. Renin secre
11. TGF-beta sig
12. Gap junction
13. GnRH signa
14. Morphine ad
15. Proteoglyca
16. MAPK signa
17. Circadian en



## Notes:

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# Further Training

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## Self-learning

- Check out <https://documentation.partek.com/display/PGS> for documentation and additional resources
- Recorded webinars available on <http://www.partek.com/webinars>

## Regional Technical Support

- Email: [support@partek.com](mailto:support@partek.com)
- Phone: +1-314-878-2329

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